Antibiotic resistance in the patient with cancer: Escalating challenges and paths forward

Amila K Nanayakkara, PhD, Division of Infectious Diseases and Geographic Medicine, Department of Medicine, University of Texas Southwestern, Dallas, Texas, USA; Helen W Boucher, MD, Division of Geographic Medicine and Infectious Diseases, Tufts Medical Center, Boston, Massachusetts, USA; Vance G Fowler, Jr, MD, MHS, Division of Infectious Diseases, Department of Medicine, Duke University Medical Center, Durham, North Carolina, USA; Amanda Jezek, Infectious Diseases Society of America, Arlington, Virginia, USA; Kevin Outterson, JD, LLM, CARB-X, Boston, Massachusetts, USA; Boston University School of Law, Boston, Massachusetts, USA and David E Greenberg, MD, Division of Infectious Diseases and Geographic Medicine, Department of Medicine, University of Texas Southwestern, Dallas, Texas, USA; Department of Microbiology, University of Texas Southwestern, Dallas, Texas, USA

Infection is the second leading cause of death in patients with cancer. Loss of efficacy in antibiotics due to antibiotic resistance in bacteria is an urgent threat against the continuing success of cancer therapy. In this review, the authors focus on recent updates on the impact of antibiotic resistance in the cancer setting, particularly on the ESKAPE pathogens (Enterococcus faecium, Staphylococcus aureus, Klebsiella pneumoniae, Acinetobacter baumannii, Pseudomonas aeruginosa, and Enterobacter spp.). This review highlights the health and financial impact of antibiotic resistance in patients with cancer. Furthermore, the authors recommend measures to control the emergence of antibiotic resistance, highlighting the risk factors associated with cancer care. A lack of data in the etiology of infections, specifically in oncology patients in United States, is identified as a concern, and the authors advocate for a centralized and specialized surveillance system for patients with cancer to predict and prevent the emergence of antibiotic resistance. Finding better ways to predict, prevent, and treat antibiotic-resistant infections will have a major positive impact on the care of those with cancer.

First published in: CA Cancer J Clin 2021;0:1-17. © 2021 The Authors. CA: A Cancer Journal for Clinicians published by Wiley Periodicals LLC on behalf of American Cancer Society. This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

acterial resistance to penicillin was encountered in patients (1) within 2 years after mass production of the antibiotic began in 1945 (2,3). Since then, the emergence of antibiotic resistance has been reported against virtually all antibiotics developed to date (4). Organizations such as the World Health Organization (WHO) and the US Centers for Disease Control and Prevention (CDC) have recognized antimicrobial resistance (AMR) as a global threat (5,6). The misuse and overuse of antibiotics is a significant driver for increasing antibiotic resistance (4,7). If the scientific community fails to manage and replenish our antibiotic supply, nearly 10 million extra deaths are predicted by 2050 due to drug-resistant infections (8-10).

In a postantibiotic era, many interventions that we currently take for granted will be threatened. These include medical advances that have occurred in general surgery (11), treatment of immunocompromised patients (12), organ transplant

Importantly, increasing levels of antibiotic resistance are already having a profound impact on the care of patients with cancer (15). End cancer as we know it is a major priority of the Biden Administration (16) as well as medical societies (17), but achieving that goal will also require action against drugresistant microbes

Infections are common in patients with cancer, and they depend upon effective antibiotics to both prevent and treat bacterial infections. Antibiotic failure in patients with cancer increases the frequency of sepsis, sepsis-related mortality, and sepsis-associated costs of care (18-23). Thus it is not surprising that oncologists have been among the first to point out the clinical impact of increasing antibacterial resistance. For example, a recent study in the United Kingdom reported that 46% of the oncologists in the United Kingdom are worried that chemotherapy as a treatment for cancer will be difficult as a result of drug-resistant infections (24). Optimizing the recipients (13), and patients with prosthetic implants (14). use of current antibiotics and discovery of novel antibiotics

Resistance t ype (Blair 2015 ²⁸)	Examples of molecular mechanisms (Bax & Griffin 2012 ²⁹)	Effected antibiotics classes (Kapoor 2017 ³⁰)	Examples of antibiotic-resistant isolates from patients with cancer (Reference)
Antibiotic	β-Lactamases	Penicillins	ESBL-producing K. pneumoniae
inactivation	Aminoglycoside- modifying enzymes	Aminoglycosides	(Zhang 2016 ³¹) ^b ESBL-producing E. coli (Cornejo-Juarez 2015 ³²) CRE K. pneumoniae (Satlin 2027 ³³)c Carbapenem-resistant A. baumannii (Bodro 2014 ³⁴) Methicillin-resistant S. aureus (MRSA) (Bodro 201434) Metallo β-lactamase-producing P. aeruginosa (Toleman 2004 ³⁵)
Antibiotic target modification	Alteration of the peptidogly can synthesis pathway	Glycopeptides	Vancomycin-resistant E. faecium (Alatorre-Fernandez et al. 2017 ⁶⁵)
et al. 2017 ⁶⁵)	Mutations in DNA gyrase	Fluoroquinolones	Fluoroquinolone-resistant clinical isolates of E. coli (Conrad 1996 ³⁷)
	Ribosomal mutations	Tetracyclines	Ci El con (Comuna 1770)
Antibiotic efflux	Overexpression of multidrug-resistant	Tetracyclines and Fluoroquinolones	Efflux pump-overexpressing K. pneumoniae and E. coli (Hamed 2018 ³⁸)
Reduced permeability of antibiotic	Downregulation or mutations in porin proteins	Penicillins	K. pneumoniae with porin deletions (Satlin 2013 ³⁹)
		Cephalosporins	

ESKAPE indicates Enterococcus faecium, Staphylococcus aureus, Klebsiella pneumoniae, Acinetobacter baumannii, Pseudomonas aeruginosa, and Enterobacter spp.

ESBLs break down and destroy some commonly used antibiotics, including penicillins and cephalosporins (Centers for Disease Control and Prevention 2019⁴⁰).
*CRE-like E. coli and K. pneumoniae develop resistance to the group of antibiotics called carbapenems (Centers for Disease Control and Prevention 2019⁴¹).

are critically important to protect patients with cancer from antibiotic-resistant infections in the future because antibiotic resistance threatens to undo much of the hard-won progress against cancer (25).

Antibiotic resistance is defined as the ability of microorganisms to survive when exposed to antibiotics that usually would kill them or prevent their growth (26). Some of the key factors contributing to antibiotic resistance are misuse of antibiotics in humans and animals, use of antibiotics in animal and food industries, lack of rapid diagnosis procedures, and the presence of antibiotics in the environment (27). Antibiotic resistance can be intrinsic or acquired due to various genetic mechanisms. We have highlighted the major mechanisms of antibiotic resistance in Table 1 (28-41). Some mechanisms can lead to antibiotic resistance in 1 or 2 classes of antibiotics. whereas others result in multidrug-resistant (MDR) isolates, which are characterized by exhibiting resistance to ≥3 different classes of antibiotics (42,43). In 2008, Rice et al designated 6 groups of bacteria (Enterococcus faecium, Staphylococcus aureus, Klebsiella pneumoniae, A. baumannii, Pseudomonas aeruginosa, and Enterobacter spp.) that were commonly associated with antibiotic resistance in the hospital environment and referred to them as ESKAPE pathogens (44). In this review, we focus on recent updates regarding antibiotic-resistant ESKAPE infections, including risk factors, antibiotic use, management, and prevention of antibiotic resistance in patients with cancer.

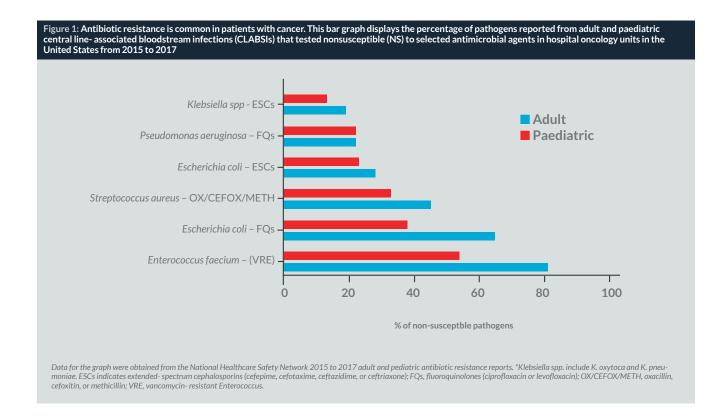
The use of antibiotics and the burden of antibiotic resistance in patients with cancer

Infections are one of the most frequent complications seen in patients with cancer (45), and a patient with cancer has a 3 times greater risk of dying from a fatal infection than a patient without cancer (46). Infections are thought to play a primary or associated role in the cause of death in approximately 50% of patients with hematological malignancies or solid tumours (47), even if drug-resistant infections are rarely recorded as the official cause of death on death certificates (48). Bacteria are the most common cause of infections in patients with cancer (47, 49). Risks of developing an infection include disruption of anatomic barriers (50), surgery, (51) chemotherapy-related and radiation-related neutropenia (52), and stem cell transplantation (53). More recently, an increased risk of infection is reportedly caused by toxicity mitigation strategies using newer immunotherapies against cancer (54-56). Under neutropenic conditions, patients with cancer are subjected to prolonged treatment of antibiotics prophylactically and empirically (57,58). However, widespread and prolonged use of broad-spectrum antibiotics to reduce mortality and morbidity from infections in patients with cancer are likely contributors to the emergence of resistance (59-61). In addition, patients with cancer are vulnerable to health care-acquired infections as a major source of antibioticresistant organisms (32,62,63). We have summarized several

POPULATION STUDIED	RESISTANT MICROORGANISMS	RISK FACTORS FOR DEVELOPMENT OF ANTIBIOTIC RESISTANCE	INTERPRETATIONS	REFERENC
SI episodes in patients with cancer	Enterococcus	Prolonged antibiotic exposure	403 Episodes of EF BSIs from 21,695 positive blood cultures	Tedim 2017
January 1995 to May 2015)	faecium (EF)		 Increase in BSIs due to EF infections observed from 2005 to 2015 	
lematologic neutropenic patients July 2009 to July 2012)	Enterococcus faecium	Previous hospitalization	 Ampicillin-resistant EF (AREfm) colonization was detected in 32 of 52 patients (61.4%) 	Sanchez-Dia 2016 ⁶⁴
		Levofloxacin extended prophylaxis	Multidrug-resistant (MDR) clones of AREfm in intestine of patients with cancer increase the development of bacterer	
SIs in patients with hematologic malignancies lanuary 2008 to December 2012)	Enterococcus faecium	Prophylactic antibiotics	58 Episodes of EF BSI episodes from a total of 15,095 blood cultures	Alatorre Fernande: 2017 ⁶
		• Vancomycin therapy during the previous 3 mo	Higher mortality was associated with vancomycin- resistant isolates	
SIs in malignant hematology and oncology	ogy Enterococcus faecium	Prior antibiotic exposure	96 Patients with EF BSIs were included in the study	Xie 2020 ⁵⁹
atients (2008-2014)			Higher 30-d mortality was associated with vancomycin-resistant isolates	
SIs in patients with hematologic malignancies		Previous hospitalization	64 Patients with PA BSIs were studied	Tofas 2020
January 2012 to December 2014)	aeruginosa (PA)	Prior use of fluoroquinolones	• 37.5% Isolates were MDR	
			PA is an important pathogen in patients who have hematologic malignancies associated with high mortality	
SIs in patients with hematologic malignancies	Pseudomonas aeruginosa		• 55 Episodes of PA bacteremia among 51 patients	Hakki 201
nd hematopoietic cell transplant recipients anuary 2012 to March 2018)			 Fluoroquinolone prophylaxis was associated with nonsusceptibility to meropenem, but not to anti-pseudomonal β-lactams or aminoglycosides 	
SIs in neutropenic patients with cancer lanuary 2006 to May 2018)	Pseudomonas aeruginosa	Prior therapy with piperacillin- tazobactam	• 1217 Episodes of BSI due to PA across 34 centers in 12 countries	Gudiol 2020 ⁶⁷
		Prior anti-pseudomonal carbapenem useFluoroquinolone prophylaxis	The rate of MDR increased significantly over the study perio	d
espiratory infections in patients with lung ancer (September 2017 to October 2018)	Klebsiella pneumoniae (KP)		KP was identified in 27 of 47 patients who had lung cancer with respiratory infection	Ding 2020
			51.4% KP isolates were MDR and the dominant strain causing lung infection in patients with lung cancer in the study	
atients who had cancer with BSIs, HAIs, and atra-abdominal infections	Klebsiella pneumoniae		• In total, 230 consecutive cases of KP infection were studied	Zhang 201
ebruary to July 2013)			• 12.6% of hypervirulent KP isolates produced extended-spectrum β-lactamase	
SIs in malignant hematology and oncology atients (January 2014 to September 2018)	Klebsiella pneumoniae	Carbapenem exposure within 30 d before the onset of BSIs	89 patients with KP bacteremia were included in the study	Liu 2019
			Carbapenem-resistant KP caused more mortality than carbapenem-susceptible KP (55.0% vs 15.9%; P=.001)	
atients with cancer (2006 to March 2015)	Methicillin-resistant Staphylococcus aureus (MRSA)		21.1% of MRSA was documented from 450 patients reported with <i>S. aureus</i> infection	Bello-Chav 2018 ⁷⁰
	,		 Protective factors for mortality included catheter remova and initiation of adequate treatment for S. aureus <48 h after positive blood cultures 	il
atients with erythrodermic cutaneous T-cell	Staphylococcus		Of 50 events, 17 (34%) were due to MRSA	Emge 202
mphoma (CTCL) (2012-2016)	aureus		The MRSA prevalence was high in patients with erythrodermic CTC	
atients with cancer une 2014 to March 2016)	Methicillin-resistant Staphylococcus aureus		120 Isolates (40 community-acquired and 80 hospital- acquired MRSA) were included in the study	Shehata 2019 ⁷
			Patients with community-acquired MRSA showed remarkable ability to acquire MDR after irradiation	
atients with cancer (July 2017 to January 2018)	cinetobacter baumanni (AB)i		48 AB isolates were recovered from 520 blood samples	Wasfi 202
			Carbapenemases were identified as the main mechanism of carbapenem resistance in AB	
ntients with cancer—outbreak initiated om a single patient (March 2011)	Acinetobacter baumannii	• HAIs	66 AB strains (62.3%) were considered infection, and 40 (37.7%) were considered colonization	Cornejo Juarez 2020 ⁶
			 Highlighted the threat that represents the transfer of colonized patients with MDR strains between institutions 	
Patients with malignant hematology January 2014 to June 2015)	Acinetobacter baumannii	Previous carbapenem exposure	accounting for 2.9% (40 of 1358) of bacteremia cases	Wang 2017
		Previous hospitalization	 Patients who had carbapenem-resistant AB infections had significantly longer hospital stays 	

studies in which ESKAPE pathogens were isolated from developing antibiotic resistance in patients with cancer. For hospital-acquired infections are the major risk factors for the percentage of central line-associated bloodstream

patients with cancer since 2015 in Table 2 (31,59-62,64-74). Figure 1, we derived data from the National Healthcare Safety These illustrate the prevalence of MDR in different ESKAPE Network (NHSN) 2015 to 2017 adult and pediatric antibiotic pathogens and highlight that prior antibiotic exposure and resistance reports (75,76). to illustrate differences between



to selected antimicrobial agents. Vancomycin resistance in E. faecium and fluoroquinolone nonsusceptibility in Escherichia coli appear to be significantly higher in adult oncology patients compared with pediatric patients.

Antibiotic resistance is related to unfavourable outcomes in patients with cancer

Antibiotic resistance leads to detrimental effects in patients with cancer, who rely on antibiotics to prevent and treat infections. Although cancer survivorship has increased with the success of modern cancer care, current therapeutic approaches continue to make these patients vulnerable to infections (77-79). A meta-analysis by Teillant et al found that, in postchemotherapy infections, 26.8% of pathogens were identified as resistant to the standard prophylactic antibiotics that had been prescribed. That study forecasted that a reduction in antibiotic efficacy of 30% to 70% would result in nearly 4,000 to 10,000 additional infections and 500 to 1,000 additional deaths per year in the United States among patients who go through chemotherapy for hematological malignancies (15).

Multiple studies demonstrate the impact of increasing resistance on outcomes in this vulnerable population (80-82). Bodro et al reported increased persistence of bacteremia (25% vs 9.7%), metastatic infection (8% vs 4%), and early casefatality rates (23% vs 11%) among patients with cancer who had

infections by ESKAPE pathogens that tested nonsusceptible compared with other bacterial pathogens. Risk factors that were associated with having an antibiotic-resistant infection included comorbidities, prior antibiotic therapy, having a urinary catheter, and a urinary tract source of infection. Those authors identified a wide variety of pathogens, including: methicillin-resistant S. aureus (MRSA), extended-spectrum β-lactamase (ESBL)-producing K. pneumoniae, carbapenemresistant A. baumannii, carbapenem-resistant and quinoloneresistant P. aeruginosa, and de-repression of chromosomal β-lactamase and ESBL-producing Enterobacter species (34).

> A study in 2015 found that 58 of 282 deaths (23%) among patients with cancer who required intensive care were caused by hospital-acquired infections. In 51 of those 58 cases (88%), an MDR pathogen was identified. The overall prevalence of MDR pathogens was nearly 40% in microorganisms collected from patients who were admitted to the intensive care unit. Of the identified MDR pathogens, 20% were caused by E. coli (94.4% of these were ESBL producers), 12% were caused by S. aureus (90.6% of these were MRSA), 12% were caused by E. faecium (18.7% were vancomycin resistant), and 6% were caused by A. baumannii (all were MDR) (32).

In 109 patients with hematological diseases who were undergoing chemotherapy, overall survival at 30 days was analyzed in those who had Gram-negative bloodstream infections (BSIs). In patients who had infections caused by MDR bacteria, survival was significantly lower compared with the survival of those who had infections caused by noninfections caused by antibiotic-resistant ESKAPE pathogens MDR isolates (85.6% vs 55.9%; P <.001) (83). In addition, numerous recent studies support the association of antibiotic resistance with unfavourable outcomes in patients with both hematological malignancies and solid tumours (84-88). The impact of resistance is not limited to the adult population. In a tertiary children's hospital from 2010 to 2014, carbapenem-resistant versus carbapenem-susceptible BSI was associated with a longer duration of bacteremia (mean, 3.8 vs 1.7 days), a higher risk for intensive care unit hospitalization (44.4% vs 10.1%), and a higher mortality rate (33% vs 5.8%) in patients with hematological malignancies and after hematopoietic stem cell transplantation (89).

Infections with antibiotic-resistant bacteria have been studied less in patients with solid tumours than in those with hematological malignancies (90). This could be because of a lower incidence of BSIs reported in solid tumours compared with hematological malignancies in neutropenic patients with cancer (91). One main difference in infections between solid and hematological malignancies is the source of infection: pneumonia and urinary tract infections were frequent among patients with solid tumours, whereas endogenous sources and catheter-related BSIs were frequent in patients with hematological malignancies (91). The risk of infection in patients with solid tumours can be increased by factors such as chemotherapy-related or radiation therapyrelated neutropenia, disruption of anatomic barriers from medical devices and surgical or diagnostic procedures, and obstruction due to primary or metastatic tumours, resulting in postobstructive pneumonia, lung abscess, or urinary tract infections. Common sites of infection in patients with solid tumours include BSIs related to neutropenia and postsurgical site infections in breast, bone, central nervous system, and skin (45). Recent epidemiologic data highlight the high prevalence of MDR pathogens in these patients (92-94). One study reported that patients older than 70 years with solid tumours had more frequent infections because of MDR organisms compared with patients younger than 70 years (87). Another study demonstrated that patients with solid tumours were more susceptible to bacteremic cholangitis caused by Enterobacteriaceae and E. faecium, highlighting the emergence of MDR as a special concern, especially in patients who have a second episode of bacteremia (95). AMR can become important even during the diagnostic evaluation of solid tumours. For example, recent literature has demonstrated complications such as increased hospitalization and death due to antibiotic-resistant infections after prostate biopsies (96,97). Extensive use of fluoroguinolone prophylaxis may be associated with an increase in resistant E. coli strains, which can result in infections after prostate biopsies (98); as a result, broad-spectrum and longer duration of prophylaxis is recommended (96,99). Importantly, targeted antibiotic

numerous recent studies support the association of antibiotic prophylaxis with prebiopsy screening has reduced the number resistance with unfavourable outcomes in patients with both of infections after the biopsy (100,101).

Cancer and antibiotic resistance also converge to worsen health disparities. Certain communities of colour in the United States, including African American, Latinx, and indigenous communities, experience higher cancer incidence and lower survival rates for many types of cancers. Many complex factors drive these disparities (102). Similarly, experts have identified many reasons to suspect a disparate impact of AMR, including differences regarding the use of prescribed and nonprescribed antibiotics, barriers to medical care, higher rates of foreign travel to regions with high AMR burden, and more likely employment in food animal production (103). Taken together, the joint epidemics of cancer and AMR can contribute significantly to persistent health inequities.

AMR and the cost of treating cancer

The decline of antibiotic effectiveness due to AMR has imposed a massive burden on health-care costs, with an increase in hospital admissions (104). Antibiotic resistance is estimated to cost nearly US\$ 20 billion in health care and US\$ 35 billion a year in lost productivity in the US economy (4,105). The cost of treating infections in patients with cancer adds a significant amount to the overall cost of cancer treatment. For example, of all-cause health-care costs during first-line chemotherapy, neutropenia-related costs accounted for 32.2% in patients with non-small lung cancer who were diagnosed with febrile neutropenia (106). On the basis of a study published with 91,560 and 16,859 cancer-related neutropenia hospitalizations among adults and children, respectively, the cost of cancerrelated neutropenia hospitalization was US\$ 24,770 per stay for adults and US\$ 26,000 per stay for children in the United States (107). Tori et al reported that the cost of treatment for an episode of febrile neutropenia after chemotherapy, on average, was from US\$ 50,000 to US\$ 60,000 in 2020 (108).

Although studies have estimated the increased cost of health care caused by AMR, the direct costs of AMR related to cancer therapy have rarely been studied. In 2004, Watters et al reported the cost associated with the treatment of patients with head and neck cancer who become colonized or infected with MRSA after major surgical procedures. Patients who were colonized or infected with MRSA had up to a 3 times more prolonged hospital stay compared with those who were not positive for MRSA. Furthermore, the authors reported that the cost of antibiotics increased by US\$ 2,470 per patient because of MRSA (109).

Strategies for preventing antibiotic resistance in patients with cancer

Prevention of infection—minimizing antibiotic usage
Antibiotic prophylaxis is a common practice for preventing

infections and infection-related complications under neutropenic conditions in patients who have cancer (110,111).

With neutropenic conditions, patients are prone to develop fever (febrile neutropenia), indicating possible infection. The mortality rate can go up to 11% in patients who have cancer with febrile neutropenia (112,113) and can be as high as 50% during severe sepsis conditions (114). According to some studies, prophylactic use of quinolones reduces the incidence of fever, probable infections, hospitalizations, (115,116) and the overall mortality rate (110,117). These gains must be balanced with observations that patients with cancer who receive prolonged antibiotic prophylaxis are at risk for developing breakthrough antibiotic-resistant infections (67,118-120).

Previous antibiotic exposure has been recognized as one of the main risk factors for AMR development in some patients with cancer (59-61,67). In fact, there remains ongoing debate in clinical oncology settings about the overall use or duration of quinolone prophylaxis in some patients with cancer because the procedure failed to reduce overall mortality and increased the emergence of resistant strains in some studies (121-124).

Minimizing infections provides an opportunity to reduce the use of antibiotics in patients with cancer who have neutropenia or those undergoing surgeries and other invasive procedures. The CDC, the American Cancer Society, and the National Comprehensive Cancer Network provide guidance to patients with cancer, caregivers, and their health-care teams to prevent infections in patients who have cancer. These include educating patients and caregivers about day-to-day good practices to prevent infections or to detect infections early (125-127).

Antibiotic or chemotherapy administration can result in gut microbiota dysbiosis, altering the diversity of bacteria (128-130). Dysbiosis in the gut microbiota can increase the risk for resistance bacteria in the microbiota (131), invasive infections, (50) post-transplant complications (such as graft-versus-host disease in those who undergo hematopoietic stem cell transplantation) (132), and reduced efficacy in patients who have cancer treated with immunotherapy (133). Monitoring gut microbiota for its composition, administering protective commensal bacteria to reduce antibiotic-resistant infections, and promoting a healthy microbiome could be promising approaches for preventing antibiotic resistance, minimizing antibiotic use, and leading to positive outcomes in these patients (134-136).

Another area of concern for patients with cancer is the recognition that there is geographical variability in antibiotic resistance. Resistance to antibiotics frequently originates in one locality, only to spread to others. For example, vancomy cinresistant *Enterococci* was identified in 1987 in Europe and, within 10 years, it represented >25% of *Enterococci* associated

with BSIs in hospitalized patients in the United States (137). A study by Arcilla et al in 2017 found that 34.3% of 1,847 travelers who were ESBL-negative before traveling from the Netherlands had acquired ESBL *Enterobacterales* during their international travel, with examples of transmission within households (138). Furthermore, medical tourists travel between health facilities in locations with different rates of antibiotic resistance, potentially disseminating resistant pathogens (139). With international travel poised to rebound after COVID-19, vulnerable groups such as patients with cancer should remain aware of infectious risks, including information on the prevalence of drug-resistant pathogens that might be present in the locations to which they travel.

Promoting the appropriate use of antibiotics among healthcare practitioners and patients will prevent the misuse and overuse of antibiotics as well as decreasing costs (140-142). Most importantly, this will allow for continued use of the existing antibiotic armamentarium (143-145). The required duration of antibiotic therapy is inexact and has been disputed in oncology settings, leading to unnecessarily extended courses of antibiotics and heterogeneity of use between practice sites (146-148). Well defined guidelines are required after comprehensive studies to establish the optimal duration of antibiotic administration to reduce antibiotic overuse in oncology settings (149). For example, vancomycin has been shown to be inappropriately prescribed as empirical treatment resulting in vancomycin resistance (150). Fever and neutropenia guidelines published by the Infectious Diseases Society of America indicate that vancomycin is not recommended as a standard part of the initial antibiotic regimen for fever and neutropenia and should be considered for specific clinical indications. Furthermore, these guidelines emphasize the importance of discontinuing vancomycin in the absence of Gram-positive organisms (151). Antibiotic deescalation and discontinuation should be considered when the patient is stabilized or the causative agent is determined to reduce overuse (152). Early discontinuation of empirical antibacterial therapy in patients with fever of unknown origin has been demonstrated to be safe, (153,154) and emerging data indicate that continuation of empirical antibiotics until absolute neutrophil count recovery could be unnecessary (155,156). De-escalating and discontinuation strategies have been successfully demonstrated in high-risk neutropenic patients who have cancer, with a significant reduction in antibiotic use (157-159).

Antibiotic stewardship to optimize antibiotic use

Antimicrobial stewardship has been defined as selection of the best antimicrobial treatment at the optimal dose and duration, resulting in the best clinical outcome for treating and on subsequent resistance (160,161).

In health-care settings, antimicrobial stewardship teams, ideally led by infectious diseases physicians in partnership with infectious diseases pharmacists, clinical microbiologists, and infection preventionists, are charged with this important initiative. Antimicrobial stewardship is especially important for patients with cancer and/or those undergoing hematopoietic stem cell transplantation, who are prone to serious infections and receive multiple courses of antimicrobial therapy during the treatment process (162). These patients may have the most potential to benefit from antibiotic stewardship because past antibiotic exposure is a critical risk factor for developing an antibiotic-resistant infection. As discussed above, patients who have cancer with antibiotic-resistant infections have worse outcomes than those who have antibiotic-susceptible infections (163). Rosa et al evaluated patient outcomes related to antibiotic stewardship in patients with febrile neutropenia, specifically, mortality in those with hematological malignancies and solid tumours. Their study indicated that adherence to antibiotic stewardship was independently associated with lower mortality (164). However, according to a review published by Pillinger et al in 2020, these patient populations are frequently excluded from studies of antibiotic stewardship, and more efforts are needed to determine the broader impact of different stewardship strategies in this vulnerable patient population (165). Nevertheless, several other studies in hospital-wide intervention programmes have demonstrated the impact of antibiotic stewardship on decreasing antibiotic resistance system wide and reducing antimicrobial expenditures (166-170). Although more data in this patient population are needed, it is reasonable to conclude that decreases in infections caused by antibioticresistant pathogens in a health-care system would translate to improved outcomes across a diverse range of patient populations. The Centers for Medicare and Medicaid Services require acute care hospitals and long-term care facilities to have antibiotic stewardship programmes in place, but their impact is uneven because many hospitals lack sufficient resources to fully implement stewardship protocols (171). Only recently has stewardship become a focus in outpatient settings, where high levels of inappropriate antibiotic prescriptions persist. Recently implemented Core Elements of Outpatient Antibiotic Stewardship by the CDC focus on a framework for antibiotic stewardship for outpatient clinicians and facilities that routinely provide antibiotic treatment (173). Increased resources will be critical to the universal adoption of stewardship, and patients at greatest risk for increased morbidity and mortality because of antibiotic-resistant

preventing infection with minimal toxicity and a minimal effect infections – such as those with cancer – have the most to gain (173).

> Other than health-care settings, it is important to focus on more general areas that contribute to the occurrence of antibiotic-resistant bacteria. Agriculture, such as the livestock and poultry industries (174), is one important area of concern. These industries consume large quantities of antibiotics to protect animals from infection and also to promote growth (175,176). According to the 2019 Summary Report on Antimicrobials Sold or Distributed for Use in Food-Producing Animals by the US Food and Drug Administration, 54% of nearly 11 metric tons of antibiotics used in animal agriculture are medically important, such as tetracyclines and penicillins (174). Antibiotic-resistant bacteria occurring in these settings can be transmitted to humans (177-179). Although no studies have been performed to correlate antibiotic resistance in farm animals and patients with cancer, it is likely that such patients could face complications because of colonization of antibioticresistant species in their intestines. Tackling antibiotic resistance will require a sustained, multi-faceted approach in numerous segments of society.

> Antibiotic-resistance surveillance systems for patients with cancer: Prediction and prevention of outbreaks

> The CDC has defined surveillance as systematic, ongoing collection, analysis, and interpretation of health data essential for planning, implementing, and evaluating public health practice integrated closely with timely dissemination to those who need the data (180). Various countries have developed their own guidelines for the surveillance of antibiotic-resistant bacteria (181-183). Surveillance of AMR involves the tracking and analysis of antibiotic-susceptibility test results in bacteria isolated from clinical samples. These results, combined with clinical and demographic data obtained from patients, enable clinicians to provide meaningful interventions to reduce the burden of antibiotic resistance (184). Surveillance data can be used for predictions. The data from surveillance, merged with other risk factors, can be used to develop prediction models for antibiotic-resistance development in clinically relevant bacterial pathogens. In 2020, Gudiol et al developed a clinical prediction model available online that could identify neutropenic patients with cancer who are at high risk of bloodstream infections because of MDR P. aeruginosa, centered on parameters such as patient age and prior antibiotic use. Although the study has not been replicated yet by other groups, the investigators reported good prediction results in patients with cancer from across 34 centres in 12 countries, indicating that the model may benefit these patients by improving the administration of specific empirical antibiotic

treatment and that it may also help optimize the effective-ness of antibiotic stewardship programmes (67). A comprehensive and predictive model of ESKAPE pathogens theoretically could be a useful tool for predicting the emergence of antibiotic resistance in oncology settings and driving the efficient utilization of antibiotics. The CDC has increased antibioticresistance surveillance in accordance with the first National Action Plan for Combating Antibiotic Resistant Bacteria, but significant gaps in our knowledge remain (185). For example, adult and pediatric antibiotic-resistance reports issued from 2015 to 2017 by the NHSN highlighted health care-associated infections from 17 adult and 8 pediatric oncology facilities only. The number of oncology facilities that reported data was relatively low compared with the total number of healthcare facilities that reported data in the NHSN (5,626 adult centers and 2,545 paediatric centres) (75,76). Furthermore, the report separately revealed the percentage of antibiotic non-susceptible pathogens recorded from oncology units, as summarized in Figure 1. A comparison of the percentage of nonsusceptible pathogens between adult and pediatric oncology units reveals higher levels of vancomycin-resistant E. faecium and fluoroquinolone-resistant E. coli. However, similar data were not found for oncology facilities from previous reports by the NHSN, so comparisons from previous years could not be made (186-189). Having chronological surveillance data on antibiotic resistance in oncology settings will be critical for tracking trends and linking rates of resistance to interventions made in these patients. Ongoing and future efforts by the CDC will help in this regard.

Future innovations in antibiotics and their impact on resistance

Althoughseveralinternational and governmental organizations have helped fund new efforts, such as CARB-X (Combating Antibiotic-Resistant Bacteria Biopharmaceutical Accelerator), to spur the development of innovative antibiotics, (190) several expert reports have warned that antibiotic business models are uniquely broken and require significant reform to bring innovative new antibacterials to patients (191-195). Because physicians frequently reserve new antibiotics as a last resort treatment for infections, this results in a low volume of sales (4). An analysis by Towse et al in 2017 estimated that the cost of developing an antibiotic is approximately US\$ 1,581 million, whereas the average annual revenue from an antibiotic's sales is roughly US\$46 million (196). This results in significant obstacles for the pharmaceutical industry to developing new antibiotics (4). A predictable return on investment for antibiotic development will likely require the support of the federal government enacting policies that could help prevent

the collapse of the antibiotic pipeline. Financial incentives for antibiotic innovation should target drugs that will provide the most clinical benefit for patients with the most significant unmet medical needs (197).

The vast majority of antibiotics used clinically are broadspectrum. Broad-spectrum antibiotics are usually active against multiple bacterial species, not just the specific pathogen that might be targeted in a particular patient scenario (198).

One major drawback of broad-spectrum antibiotics is the development of AMR not only in pathogenic bacteria but also in the non-pathogenic commensal bacteria that comprise the normal microbiome (199). The development of narrowspectrum antibiotics is considered an attractive approach to overcoming antibiotic-resistant bacterial infections because more specific antibiotics can reduce the selection pressure in non-targeted pathogens (200,201). Examples of experimental narrow-spectrum antibiotics for ESKAPE pathogens include bacteriophages, (202,203) monoclonal antibodies (204), bacteriocins (205,206), and antisense molecules, such peptide-conjugated phosphorodiamidate morpholino oligomers (207-209). Bacteriophages are bacterial viruses that infect bacterial cells, which can cause the bacterium to lyse (210). Bacteriophages are specific for bacteria and selectively attach to specific receptors on the surface of the host cell (211). Similar to phages, human monoclonal antibodies also can be developed for specific bacteria and can be targeted by the immune system (212,213). Bacteriocins are peptides of different sizes produced by various bacteria that exhibit bactericidal activity against other bacteria (205,214). Bacteriocins bind various receptors on the surface of the target bacteria to trigger bactericidal effects (215). Phosphorodiamidate morpholino oligomers are designed to target mRNA and block translation of the gene of interest (207). Continuing advances in the rapid identification of pathogens will enable the opportunity of using narrow-spectrum antibiotics. Recent developments in diagnostic tests, such as next-generation sequencing, (36,216-218) matrix-assisted laser desorption ionization-time-offlight mass spectrometry (219), and rapid antigen testing, (220) have made the prospect of pathogen-specific therapy a viable strategy. Additional policies are needed to strengthen diagnostic innovation and clinical integration of diagnostics, including better outcomes studies to inform clinical use and justify appropriate reimbursement (221). Recently, the Infectious Diseases Society of America issued new guidelines to treat antimicrobial-resistant, Gram-negative infections focusing on the efficiency of different antibiotics according to the etiology of the infection. These guidelines provide preferred or alternative antibiotic treatment options with dosages for ESBL-producing Enterobacterales, carbapenemresistant *Enterobacterales*, and difficult-to-treat *P. aeruginosa* according to the source of infection (222).

Conclusion

Drug-resistant infections are growing in number and cost and significantly threaten our ability to care for patients with cancer. The cancer community – patients, loved ones, clinicians, and scientists – have successfully advocated for significant investments in research and public health strategies to prevent cancer and increase therapeutic options, with the goal of saving and extending lives (223). Because antibiotic resistance threatens to undo much of this hard-won progress, cancer advocates should consider focusing their considerable political power on this public health crisis. Cancer and infectious diseases experts must unite to drive the federal policy changes necessary to prevent, diagnose, and treat drug-resistant infections and to protect the gains that have been made against cancer over the past few decades.

Disclosures

No specific funding was disclosed. Helen W Boucher reports honoraria as editor of Infectious Diseases Clinics of North America, Antimicrobial Agents and Chemotherapy, and the Sanford Guide. Vance G Fowler, Jr, reports personal fees from Novartis, Novadigm, Durata, Debiopharm, Genentech, Achaogen, Affinium, Medicines

Company, Cerexa, Tetraphase, Trius, MedImmune, Bayer, Theravance, Basilea, Affinergy, Janssen, xBiotech, Contrafect, Regeneron, Basilea, Destiny, Amphliphi Biosciences, Integrated Biotherapeutics, C3J, Armata, Valanbio, Akagera, and Aridi; grants from the National Institutes of Health, MedImmune, Allergan, Pfizer, Advanced Liquid Logics, Theravance, Novartis, Merck, Medical Biosurfaces, Locus, Affinergy, Contrafect, Karius, Genentech, Regeneron, Basilea, and Janssen; royalties from UpToDate; stock options in Valanbio; a patent pending in sepsis diagnostics; educational fees from Green Cross, Cubist, Cerexa, Durata, Theravance, and Debiopharm; and an editor's stipend from the Infectious Diseases Society of America all outside the submitted work. Kevin Outterson reports grants from the US Biomedical Advanced Research and Development Authority, the Wellcome Trust (United Kingdom), the Global Antimicrobial Resistance Innovation Fund (United Kingdom), the Federal Ministry of Education and Research (Germany), and the Bill and Melinda Gates Foundation outside the submitted work. David E. Greenberg reports grants from the National Institutes of Health, the US Department of Defense, and Shionogi; personal fees from MDC Associates; patents relating to antisense molecules and alternating magnetic fields; and serves as the Chief Medical Officer and founder of Solenic Medical, for which he has stock options and receives consultant fees, all outside the submitted work. Amila K. Nanayakkara and Amanda Jezek had no disclosures.

References

- Spink WW, Ferris V. Penicillin-resistant staphylococci: mechanisms involved in the development of resistance. J Clin Invest. 1947;26:379-393. doi:10.1172/ JCI10 1820
- Aminov RI. A brief history of the antibiotic era: lessons learned and challenges for the future. Front Microbiol. 2010;1:134. doi:10.3389/fmicb.2010.00134
- 3. Hamilton KW. Miracle Cure: The Creation of Antibiotics and the Birth of Modern Medicine. *Emerging Infect Dis.* 2019;25:196. doi:10.3201/eid25 01.181184
- 4. Ventola CL. The antibiotic resistance crisis: part 1: causes and threats. P T. 2015;40:277-283
- 5. Centers for Disease Control and Prevention. Antibiotic Resistance Threats in the United States, 2019 (2019 AR Threats Report). Accessed February 25, 2021. cdc.gov/ drugresistance/biggest-threats.html.
- World Health Organization. Global action plan on antimicrobial resistance. 2021 Accessed February 25, 2021. who.int/publications/i/item/97892 41509763
- 7. Goossens H, Ferech M, Vander Stichele R, Elseviers M, ESAC Project Group. Outpatient antibiotic use in Europe and association with resistance: a cross- national database study. Lancet. 2005;365:579-587. doi:10.1016/S0140-6736(05)17907-0
- 8. O'Neill J. Review on Antimicrobial Resistance. Antimicrobial Resistance: Tackling a Crisis for the Health and Wealth of Nations. *Review on Antimicrobial Resistance*; 2014. Accessed February 25. 2021. amr-review.org/sites/default/files/AMR%20Rev iew%20Paper%20%20Tackling %20a%20crisis%20for%20the%20health%20and%20wealth%20of%20 nations 1.pdf
- de Kraker ME, Stewardson AJ, Harbarth S. Will 10 million people die a year due to antimicrobial resistance by 2050? PLoS Med. 2016;13:e1002184. doi:10.1371/journ al.pmed.1002184
- Hollis A, Ahmed Z. Preserving antibiotics, rationally. N Engl J Med. 2013;369:2474-2476. doi:10.1056/NEJMp 1311479
- Cohen ME, Salmasian H, Li J, et al. Surgical antibiotic prophylaxis and risk for postoperative antibiotic-resistant in- fections. J Am Coll Surg. 2017;225:631-638.e3. doi:10.1016/j.jamco llsurg.2017.08.010
- 12. DeNegre AA, Ndeffo Mbah ML, Myers K, Fefferman NH. Emergence of antibiotic resistance in immunocompromised host populations: a case study of emerging antibiotic resistant tuberculosis in AIDS patients. PLoS One. 2019;14:e0212969. doi:10.1371/journal.pone.0212969
- Bartoletti M, Giannella M, Tedeschi S, Viale P. Multidrug- resistant bacterial infections in solid organ transplant candidates and recipients. *Infect Dis Clin North Am.* 2018;32:551-580. doi:10.1016/j.idc. 2018.04.004
- 14. Ravi S, Zhu M, Luey C, Young SW. Antibiotic resistance in early periprosthetic joint infection. ANZ J Surg. 2016;86:1014-1018. doi:10.1111/ans.13720
- Teillant A, Gandra S, Barter D, Morgan DJ, Laxminarayan R. Potential burden of antibiotic resistance on surgery and cancer chemotherapy antibiotic prophylaxis in the USA: a literature review and modelling study. *Lancet Infect Dis.* 2015;15:1429-1437. doi:10.1016/S1473-3099(15)00270-4
- Gourd E. Biden determined to "end cancer as we know it." Lancet Oncol. 2021;22:759. doi:10.1016/S1470 - 2045(21)00282 - 5
- 17. Ma J, Jemal A, Fedewa SA, et al. The American Cancer Society 2035 challenge goal on cancer mortality reduction. CA Cancer J Clin. 2019;69:351-362. doi:10.3322/caac.21564
- Vazquez- Lopez R, Rivero Rojas O, Ibarra Moreno A, et al. Antibiotic- resistant septicemia in pediatric oncology patients associated with post-therapeutic neutropenic fever. Antibiotics (Basel). 2019;8:106. doi:10.3390/antibiotics8030106
- Williams MD, Braun LA, Cooper LM, et al. Hospitalized cancer patients with severe sepsis: analysis of incidence, mortality, and associated costs of care. Crit Care. 2004;8:R291- R298. doi:10.1186/cc2893
- Montassier E, Batard E, Gastinne T, Potel G, de La Cochetiere MF. Recent changes in bacteremia in patients with cancer: a systematic review of epidemiology and antibiotic resistance. Eur J Clin Microbiol Infect Dis. 2013;32:841-850. doi:10.1007/s1009 6-013-1819-7
- 21. Moghnieh R, Estaitieh N, Mugharbil A, et al. Third generation cephalosporin resistant Enterobacteriaceae and multidrug resistant Gram-negative bacteria causing bacteremia in febrile neutropenia adult cancer patients in Lebanon, broad spectrum antibiotics use as a major risk factor, and correlation with poor prognosis. Front Cell Infect Microbiol. 2015;5:11. doi:10.3389/fcimb.2015.00011
- 22. Trecarichi EM, Tumbarello M. Antimicrobial- resistant Gram- negative bacteria in febrile neutropenic patients with cancer: current epidemiology and clinical impact. Curr Opin Infect Dis. 2014;27:200-210. doi:10.1097/QCO.000000000000000038
- 23. Thorpe KE, Joski P, Johnston KJ. Antibiotic- resistant infection treatment costs have doubled since 2002, now exceeding \$2 billion annually. *Health Aff* (Millwood). 2018;37:662-669. doi:10.1377/hltha ff.2017.1153
- 24. The Longitude Prize. Effectiveness of Cancer Treatments Threatened by Rising Antibiotic Resistance. Accessed November 24, 2020. longitude prize.org/resources/effectiveness-cancer-treatments-threatened-rising-antibiotic-resistance
- 25. Lee CR, Cho IH, Jeong BC, Lee SH. Strategies to minimize antibiotic resistance. *Int J Environ Res Public Health*. 2013;10:4274-4305. doi:10.3390/ijerp h1009 4274
- Li B, Webster TJ. Bacteria antibiotic resistance: new challenges and opportunities for implant- associated orthopedic infections. J Orthop Res. 2018;36:22-32. doi:10.1002/ jor.23656

- Giacomini E, Perrone V, Alessandrini D, Paoli D, Nappi C, Degli Esposti L. Evidence
 of antibiotic resistance from population-based studies: a narrative review. *Infect Drug Resist*. 2021;14:849-858. doi:10.2147/IDR.S289741
- Blair JM, Webber MA, Baylay AJ, Ogbolu DO, Piddock LJ. Molecular mechanisms of antibiotic resistance. Nat Rev Microbiol. 2015;13:42-51. doi:10.1038/nrmic ro3380
- 29. Bax R, Griffin D. Introduction to antibiotic resistance. Handb Exp Pharmacol. 2012;211:1- 12. doi:10.1007/978- 3- 642- 28951 4_1
- Kapoor G, Saigal S, Elongavan A. Action and resistance mechanisms of antibiotics: a guide for clinicians. J Anaesthesiol Clin Pharmacol. 2017;33:300-305. doi:10.4103/joacp. JOACP 349 15
- 31. Zhang Y, Zhao C, Wang Q, et al. High prevalence of hypervirulent Klebsiella pneumoniae infection in China: geographic distribution, clinical characteristics, and antimicrobial resistance. Antimicrob Agents Chemother. 2016;60:6115-6120. doi:10.1128/AAC.01127-16
- 32. Cornejo- Juarez P, Vilar-Compte D, Perez- Jimenez C, Namendys-Silva SA, Sandoval-Hernandez S, Volkow- Fernandez P. The impact of hospital- acquired infections with multidrug-resistant bacteria in an oncology intensive care unit. *Int J Infect Dis.* 2015;31:31-34. doi:10.1016/j. ijid.2014.12.022
- 33. Satlin MJ, Chen L, Patel G, et al. Multicenter clinical and molecular epidemiological analysis of bacteremia due to carbapenem-resistant Enterobacteriaceae (CRE) in the CRE epicenter of the United States. Antimicrob Agents Chemother. 2017;61:e02349-16. doi:10.1128/AAC.02349-16
- Bodro M, Gudiol C, Garcia-Vidal C, et al. Epidemiology, antibiotic therapy and outcomes
 of bacteremia caused by drug- resistant ESKAPE pathogens in cancer patients. Support
 Care Cancer. 2014;22:603-610. doi:10.1007/s0052 0-013-2012-3
- 35. Toleman MA, Rolston K, Jones RN, Walsh TR. blaVIM- 7, an evolutionarily distinct metallo-beta- lactamase gene in a Pseudomonas aeruginosa isolate from the United States. Antimicrob Agents Chemother. 2004;48:329-332. doi:10.1128/AAC.48.1.329-332.2004
- Kim J, Greenberg DE, Pifer R, et al. VAMPr: VAriant Mapping and Prediction of antibiotic resistance via explainable features and machine learning. PLoS Comput Biol. 2020;16:e1007511. doi:10.1371/journal.pcbi.1007511
- 37. Conrad S, Oethinger M, Kaifel K, Klotz G, Marre R, Kern WV. gyrA mutations in high-level fluoroquinolone- resistant clinical isolates of Escherichia coli. J Antimicrob Chemother. 1996;38:443-455. doi:10.1093/jac/38.3.443
- Hamed SM, Elkhatib WF, El- Mahallawy HA, Helmy MM, Ashour MS, Aboshanab KMA. Multiple mechanisms contributing to ciprofloxacin resistance among Gram negative bacteria causing infections to cancer patients. Sci Rep. 2018;8:12268.
- doi:10.1038/s41598-018-30756-4
- 39. Satlin MJ, Calfee DP, Chen L, et al. Emergence of carbapenem- resistant
 Enterobacteriaceae as causes of bloodstream infections in patients with hematologic
 malignancies. *Leuk Lymphoma*. 2013;54:799-806. doi:10.3109/10428194.2012.723210
- 40. Centers for Disease Control and Prevention (CDC). ESBL- Producing Enterobacterales in Healthcare Settings. CDC; 2019. Accessed July 19, 2021. cdc.gov/hai/ organ isms/
- Centers for Disease Control and Prevention (CDC). Carbapenem-Resistant Enterobacterales (CRE). CDC; 2019. Accessed July 19, 2021. cdc.gov/hai/organisms/cre/index.html
- 42. Centers for Disease Control and Prevention. Antibiotic Resistance & Patient Safety Portal. Accessed May 28, 2021. arpsp.cdc.gov/profile/antibiotic-resistance ?tab=antibiotic-resistance
- 43. Centers for Disease Control and Prevention. Glossary of Terms Related to Antibiotic Resistance. Accessed May 28, 2021. cdc.gov/narms/resources/glossary.html
- 44. Rice LB. Federal funding for the study of antimicrobial resistance in nosocomial pathogens: no ESKAPE. *J Infect Dis.* 2008;197:1079-1081. doi:10.1086/533452
- 45. Rolston KV. Infections in cancer patients with solid tumors: a review. *Infect Dis Ther.* 2017;6:69-83. doi:10.1007/s4012 1- 017-0146-1
- 46. Zheng Y, Chen Y, Yu K, et al. Fatal infections among cancer patients: a population-based study in the United States. *Infect Dis Ther*. 2021;10:871-895. doi:10.1007/s40121-021-00433-7
- 47. Zembower TR. Epidemiology of infections in cancer patients. *Cancer Treat Res.* 2014;161:43-89. doi:10.1007/978-3-319-04220-6_2
- 48. O'Dowd A. Death certificates should record antimicrobial resistance as cause of deaths, says CMO. BMJ. 2018;362:k3832. doi:10.1136/k3832
- O'Connor D, Bate J, Wade R, et al. Infection-related mortality in children with acute lymphoblastic leukemia: an analysis of infectious deaths on UKALL2003. *Blood*. 2014;124:1056-1061. doi:10.1182/blood-2014-03-560847
- 50. Taur Y, Pamer EG. Microbiome mediation of infections in the cancer setting. *Genome Med.* 2016;8:40. doi:10.1186/s1307 3-016-0306-z
- 51. Park SY, Kim MS, Eom JS, Lee JS, Rho YS. Risk factors and etiology of surgical site infection after radical neck dissection in patients with head and neck cancer. Korean J Intern Med. 2016;31:162-169. doi:10.3904/kiim.2016.31.1.162
- Rapoport BL. Management of the cancer patient with infection and neutropenia. Semin Oncol. 2011;38:424-430. doi:10.1053/j.semin oncol.2011.03.013
- 53. Alonso CD, Marr KA. Clostridium difficile infection among hematopoietic stem

- cell transplant recipients: beyond colitis. *Curr Opin Infect Dis.* 2013;26:326-331. doi:10.1097/QCO.0b013e3283630c4c
- 54. Kyi C, Hellmann MD, Wolchok JD, Chapman PB, Postow MA. Opportunistic infections in patients treated with immunotherapy for cancer. *J Immunother Cancer*. 2014;2:19. doi:10.1186/2051-1426-2-19
- 55. Hotchkiss RS, Monneret G, Payen D. Sepsis- induced immunosuppression: from cellular dysfunctions to immunotherapy. *Nat Rev Immunol*. 2013;13:862-874. doi:10.1038/pri3552
- 56. Del Castillo M, Romero FA, Arguello E, Kyi C, Postow MA, Redelman-Sidi G. The spectrum of serious infections among patients receiving immune checkpoint blockade for the treatment of melanoma. Clin Infect Dis. 2016;63:1490-1493. doi:10.1093/cid/ ciw539
- Lustberg MB. Management of neutropenia in cancer patients. Clin Adv Hematol Oncol. 2012;10:825-826.
- 58. Scheler M, Lehrnbecher T, Groll AH, et al. Management of children with fever and neutropenia: results of a survey in 51 pediatric cancer centers in Germany, Austria, and Switzerland. *Infection*. 2020;48:607-618. doi:10.1007/s15010-020-01462-z
- 59. Xie O, Slavin MA, Teh BW, Bajel A, Douglas AP, Worth LJ. Epidemiology, treatment and outcomes of bloodstream infection due to vancomycin-resistant enterococci in cancer patients in a vanB endemic setting. *BMC Infect Dis.* 2020;20:228. doi:10.1186/s1287 9-020-04952-5
- 60. Tedim AP, Ruiz-Garbajosa P, Rodriguez MC, et al. Long-term clonal dynamics of Enterococcus faecium strains causing bloodstream infections (1995-2015) in Spain. J Antimicrob Chemother. 2017;72:48-55. doi:10.1093/jac/dkw366
- 61. Tofas P, Samarkos M, Piperaki ET, et al. Pseudomonas aeruginosa bacteraemia in patients with hematologic malignancies: risk factors, treatment and outcome. *Diagn Microbiol Infect Dis.* 2017;88:335-341. doi:10.1016/j.diagm icrobio.2017.05.00 3
- 62. Cornejo- Juarez P, Cevallos MA, Castro- Jaimes S, et al. High mortality in an outbreak of multidrug resistant Acinetobacter baumannii infection introduced to an oncological hospital by a patient transferred from a general hospital. PLoS One. 2020;15:e0234684. doi:10.1371/journ al.pone.0234684
- 63. Cornejo-Juarez P, Vilar- Compte D, Garcia-Horton A, Lopez- Velazquez M, Namendys-Silva S, Volkow- Fernandez P. Hospital-acquired infections at an oncolog- ical intensive care cancer unit: differences between solid and hematological cancer patients. *BMC Infect Dis.* 2016;16:274. doi:10.1186/s1287 9-016 - 1592-1
- 64. Sanchez-Diaz AM, Cuartero C, Rodriguez JD, et al. The rise of ampicillinresistant Enterococcus faecium high-risk clones as a frequent intestinal colonizer in oncohaematological neutropenic patients on levofloxacin prophylaxis: a risk for bacteraemia? Clin Microbiol Infect. 2016;22:59.e1-59.e8. doi:10.1016/j.cmi. 2015.08.008
- 65. Alatorre- Fernandez P, Mayoral-Teran C, Velazquez- Acosta C, et al. A polyclonal outbreak of bloodstream infections by Enterococcus faecium in patients with hematologic malignancies. Am J Infect Control. 2017;45:260-266. doi:10.1016/j. ajic.2016.10.002
- 66. Hakki M, Humphries RM, Hemarajata P, et al. Fluoroquinolone prophylaxis selects for meropenem-nonsusceptible Pseudomonas aeruginosa in patients with hematologic malignancies and hematopoietic cell transplant recipients. Clin Infect Dis. 2019;68:2045-2052. doi:10.1093/cid/civ825
- 67. Gudiol C, Albasanz- Puig A, Laporte- Amargos J, et al. Clinical predictive model of multidrug resistance in neutropenic cancer patients with bloodstream infection due to Pseudomonas aeruginosa. *Antimicrob Agents Chemother*. 2020;64:e02494- 19. doi:10.1128/AAC.02494- 19
- 68. Ding L, Yang Z, Lu J, et al. Characterization of phenotypic and genotypic traits of Klebsiella pneumoniae from lung cancer patients with respiratory infection. *Infect Drug Resist.* 2020;13:237-245. doi:10.2147/IDR.5229085
- 69. Liu J, Wang H, Huang Z, et al. Risk factors and outcomes for carbapenem-resistant Klebsiella pneumoniae bacteremia in onco- hematological patients. *J Infect Dev Ctries*. 2019;13:357-364. doi:10.3855/jidc.11189
- Bello- Chavolla OY, Bahena-Lopez JP, Garciadiego- Fosass P, et al. Bloodstream infection caused by S. aureus in patients with cancer: a 10- year longitudinal singlecenter study. Support Care Cancer. 2018;26:4057-4065. doi:10.1007/s0052 0-018-4275-1
- 71. Emge DA, Bassett RL, Duvic M, Huen AO. Methicillin- resistant Staphylococcus aureus (MRSA) is an important pathogen in erythrodermic cutaneous T- cell lymphoma (CTCL) patients. Arch Dermatol Res. 2020;312:283-288. doi:10.1007/
- s0040 3- 019- 02015 7
- 72. Shehata MMK, Radwan SM, Ali SAM. Effects of gamma-irradiation on anti- biotic resistance and diagnostic molecular markers of methicillin-resistant Staphylococcus aureus in Egyptian cancer patients. Int J Radiat Biol. 2019;95:1728- 1743. doi:10.1080/09553002.2019.1664785
- 73. Wasfi R, Rasslan F, Hassan SS, Ashour HM, Abd El-Rahman OA. Co-existence of carbapenemase-encoding genes in Acinetobacter baumannii from cancer patients. *Infect Dis Ther.* 2021;10:291-305. doi:10.1007/s40121-020-00369-4
- 74. Wang X, Zhang L, Sun A, et al. Acinetobacter baumannii bacteraemia in patients with haematological malignancy: a multicentre retrospective study from the Infection

- Working Party of Jiangsu Society of Hematology. Eur J Clin Microbiol Infect Dis. 2017;36:1073-1081. doi:10.1007/s1009 6-016-2895-2
- 75. Weiner- Lastinger LM, Abner S, Benin AL, et al. Antimicrobial-resistant pathogens associated with pediatric healthcare-associated infections: sum-mary of data reported to the National Healthcare Safety Network, 2015- 2017. Infect Control Hosp Epidemiol. 2020;41:19-30. doi:10.1017/ice.2019.297
- 76. Weiner-Lastinger LM, Abner S, Edwards JR, et al. Antimicrobial-resistant patho-gens associated with adult healthcare-associated infections: summary of data reported to the National Healthcare Safety Network, 2015-2017. Infect Control Hosp Epidemiol. 2020;41:1-18. doi:10.1017/ice.2019.296
- 77. Allam O, Park KE, Chandler L, et al. The impact of radiation on lymphedema: a review of the literature. *Gland Surg.* 2020;9:596-
- 602. doi:10.21037/gs.2020.03.20
- Paskett ED, Stark N. Lymphedema: knowledge, treatment, and impact among breast cancer survivors. Breast J. 2000;6:373-378. doi:10.1046/j.1524-4741.2000.99072.x
- 79. Moore JX, Akinyemiju T, Bartolucci A, Wang HE, Waterbor J, Griffin R. A prospective study of cancer survivors and risk of sepsis within the REGARDS cohort. *Cancer Epidemiol.* 2018;55:30-38. doi:10.1016/j.canep.2018.05.001
- 80. Johnson K, Boucher HW. Editorial commentary: imminent challenges: carbapenemresistant Enterobacteriaceae in transplant recipients and patients with hematologic malignancy. Clin Infect Dis. 2014;58:1284-1286. doi:10.1093/cid/ciu056
- 81. Zhao Y, Lin Q, Liu L, et al. Risk factors and outcomes of antibiotic-resistant Pseudomonas aeruginosa bloodstream infection in adult patients with acute leukemia. Clin Infect Dis. 2020;71(suppl 4):S386-S393. doi:10.1093/cid/ciaa1522
- 82. Pouch SM, Satlin MJ. Carbapenem-resistant Enterobacteriaceae in special populations: solid organ transplant recipients, stem cell transplant recipients, and patients with hematologic malignancies. *Virulence*. 2017;8:391-402. doi:10.1080/21505 594.2016.1213472
- 83. Scheich S, Weber S, Reinheimer C, et al. Bloodstream infections with gram- negative organisms and the impact of multidrug resistance in patients with hematological malignancies. Ann Hematol. 2018;97:2225-2234. doi:10.1007/s0027
- 7-018-3423-
- 84. Cattaneo C, Di Blasi R, Skert C, et al. Bloodstream infections in haematological cancer patients colonized by multidrug- resistant bacteria. *Ann Hematol.* 2018;97:1717-1726. doi:10.1007/s00277-018-3341-6
- 85. Levene I, Castagnola E, Haeusler GM. Antibiotic- resistant Gram- negative blood stream infections in children with cancer: a review of epidemiology, risk factors, and outcome. *Pediatr Infect Dis J.* 2018;37:495-498. doi:10.1097/INF.000000000000001938
- 86. Marin M, Gudiol C, Garcia- Vidal C, Ardanuy C, Carratala J. Bloodstream infections in patients with solid tumors: epidemiology, antibiotic therapy, and outcomes in 528 episodes in a single cancer center. *Medicine* (Baltimore). 2014;93:143-149. doi:10.1097/MD.000000000000000000
- 87. Antonio M, Gudiol C, Royo-Cebrecos C, Grillo S, Ardanuy C, Carratala J. Current etiology, clinical features and outcomes of bacteremia in older patients with solid tumors. *J Geriatr Oncol.* 2019;10:246-251. doi:10.1016/j.jgo.2018.06.011
- 88. Zhang Y, Zheng Y, Dong F, et al. Epidemiology of febrile neutropenia episodes with Gram- negative bacteria infection in patients who have undergone chemotherapy for hematologic malignancies: a retrospective study of 10 years' data from a single center. Infect Drug Resist. 2020;13:903-910. doi:10.2147/IDR.S241263
- 89. Vinker- Shuster M, Stepensky P, Temper V, Shayovitz V, Masarwa R, Averbuch D. Gram-negative bacteremia in chil- dren with hematologic malignancies and following hematopoietic stem cell transplantation: epidemiology, resistance, and outcome. *J Pediatr Hematol Oncol*. 2019;41:e493- e498. doi:10.1097/ MPH.00000 00000 001556
- 90. Gudiol C, Aguado JM, Carratala J. Bloodstream infections in patients with solid tumors. Virulence. 2016;7:298-308. doi:10.1080/21505 594.2016.1141161
- Marin M, Gudiol C, Ardanuy C, et al. Bloodstream infections in neutropenic patients with cancer: differences between patients with haematological malignancies and solid tumours. J Infect. 2014;69:417-423. doi:10.1016/j.jinf.2014.05.018
- 92. Shrestha G, Wei X, Hann K, et al. Bacterial profile and antibiotic resistance among cancer patients with urinary tract infection in a national tertiary cancer hospital of Nepal. *Trop Med Infect Dis.* 2021;6:49. doi:10.3390/tropi calme d6020049
- Fentie A, Wondimeneh Y, Balcha A, Amsalu A, Adankie BT. Bacterial profile, antibiotic resistance pattern and associated factors among cancer patients at University of Gondar Hospital, Northwest Ethiopia. *Infect Drug Resist.* 2018;11:2169-2178. doi:10.2147/IDR. 5192322
- Islas- Munoz B, Volkow- Fernandez P, Ibanes- Gutierrez C, Villamar- Ramirez A, Vilar-Compte D, Cornejo- Juarez P. Bloodstream infections in cancer patients. Risk factors associated with mortality. Int J Infect Dis. 2018;71:59-64. doi:10.1016/i.iiid.2018.03.022
- 95. Royo- Cebrecos C, Gudiol C, Garcia J, et al. Characteristics, aetiology, antimicrobial resistance and outcomes of bacteraemic cholangitis in patients with solid tumours: a prospective cohort study. J Infect. 2017;74:172- 178. doi:10.1016/j. jinf.2016.10.008
- Boehm K, Siegel FP, Schneidewind L, et al. Antibiotic prophylaxis in prostate biopsies: contemporary practice patterns in Germany. Front Surg. 2018;5:2. doi:10.3389/fsurg.2018.00002

- 97. Johansen TEB, Zahl PH, Baco E, et al. Antibiotic resistance, hospitalizations, and mortality related to prostate biopsy: first report from the Norwegian Patient Registry. World J Urol. 2020;38:17-26. doi:10.1007/s00345-019-02837-0
- 98. Derin O, Fonseca L, Sanchez- Salas R, Roberts MJ. Infectious complications of prostate biopsy: winning battles but not war. World J Urol. 2020;38:2743-2753. doi:10.1007/s00345-020-03112-399. Sieczkowski M, Gibas A, Wasik A, et al. Drug-eluting biopsy needle as a novel strategy for antimicrobial prophylaxis in transrectal prostate biopsy. *Technol Cancer Res Treat*. 2017;16:1038-1043. doi:10.1177/1533034617722080
- 100. Ryu JW, Jung SI, Ahn JH, et al. Povidone- iodine rectal cleansing and targeted antimicrobial prophylaxis using rectal swab cultures in men undergoing transrectal ultrasound- guided prostate biopsy are associated with reduced incidence of postoperative infectious complications. *Int Urol Nephrol.* 2016;48:1763-1770. doi:10.1007/s11255-016-1394-9
- 101. Cussans A, Somani BK, Basarab A, Dudderidge TJ. The role of targeted prophylactic antimicrobial therapy before transrectal ultrasonography-guided prostate biopsy in reducing infection rates: a systematic review. BJU Int. 2016;117:725-731. doi:10.1111/ biu.13402
- 102. Zavala VA, Bracci PM, Carethers JM, et al. Cancer health disparities in racial/ethnic minorities in the United States. *Br J Cancer*. 2021;124:315-332. doi:10.1038/s41416-020-01038-6
- 103. Nadimpalli ML, Chan CW, Doron S. Antibiotic resistance: a call to action to prevent the next epidemic of inequality. *Nat Med.* 2021;27:187-188. doi:10.1038/s41591-020-01201-9
- 104. Dadgostar P. Antimicrobial resistance: implications and costs. *Infect Drug Resist*. 2019;12:3903-3910. doi:10.2147/ IDR.S234610
- 105. Nelson RE, Hatfield KM, Wolford H, et al. National estimates of healthcare costs associated with multidrug-resistant bacterial infections among hospitalized patients in the United States. Clin Infect Dis. 2021;72(suppl 1):S17-S26. doi:10.1093/cid/ciaa1581
- 106. Stokes ME, Muehlenbein CE, Marciniak MD, et al. Neutropenia- related costs in patients treated with first- line chemotherapy for advanced non-small cell lung cancer. *J Manag Care Pharm.* 2009;15:669-682. doi:10.18553/jmcp. 2009.15.8.669
- 107. Tai E, Guy GP, Dunbar A, Richardson LC. Cost of cancer- related neutropenia or fever hospitalizations, United States, 2012. J Oncol Pract. 2017;13:e552-e561. doi:10.1200/JOP.2016.019588
- 108. Tori K, Tansarli GS, Parente DM, Kalligeros M, Ziakas PD, Mylonakis E. The cost-effectiveness of empirical antibiotic treatments for high-risk febrile neutropenic patients: a decision analytic model. *Medicine* (Baltimore). 2020;99:e20022. doi:10.1097/MD.000000000000222
- 109. Watters K, O'Dwyer TP, Rowley H. Cost and morbidity of MRSA in head and neck cancer patients: what are the consequences? J Laryngol Otol. 2004;118:694-699. doi:10.1258/0022215042244732
- 110. Gafter- Gvili A, Fraser A, Paul M, et al. Antibiotic prophylaxis for bacterial infections in afebrile neutropenic patients following chemotherapy. *Cochrane Database Syst Rev.* 2012;1:CD004386. doi:10.1002/14651858.CD004386.pub3
- 111. Ziegler M, Landsburg D, Pegues D, et al. Fluoroquinolone prophylaxis is highly effective for the prevention of central line- associated bloodstream infections in autologous stem cell transplant patients. *Biol Blood Marrow Transplant*. 2019;25:1004-1010. doi:10.1016/j.bbmt. 2018.11.023
- 112. Carmona- Bayonas A, Jimenez- Fonseca P, Virizuela Echaburu J, et al. Prediction of serious complications in patients with seemingly stable febrile neutropenia: validation of the Clinical Index of Stable Febrile Neutropenia in a prospective cohort of patients from the FINITE study. J Clin Oncol. 2015;33:465-471. doi:10.1200/JCO.2014.57.2347
- 113. Kuderer NM, Dale DC, Crawford J, Cosler LE, Lyman GH. Mortality, morbidity, and cost associated with febrile neutropenia in adult cancer patients. Cancer. 2006;106:2258-2266. doi:10.1002/cncr.21847
- 114. Legrand M, Max A, Peigne V, et al. Survival in neutropenic patients with severe sepsis or septic shock. *Crit Care Med*. 2012;40:43-49. doi:10.1097/CCM. 0b013 e3182 2b50c2
- 115. Bucaneve G, Micozzi A, Menichetti F, et al. Levofloxacin to prevent bacterial infection in patients with cancer and neutropenia. N Engl J Med. 2005;353:977-987. doi:10.1056/NEJMo a044097
- 116. Cullen M, Steven N, Billingham L, et al. Antibacterial prophylaxis after chemotherapy for solid tumors and lymphomas. N Engl J Med. 2005;353:988-998. doi:10.1056/NEJMoa050078
- 117. Gafter- Gvili A, Fraser A, Paul M, Leibovici L. Meta- analysis: antibiotic prophylaxis reduces mortality in neutropenic patients. Ann Intern Med. 2005;142 (12 pt 1):979-995. doi:10.7326/0003-4819-142-12_part_1-20050 6210-00008
- 118. Razonable RR, Litzow MR, Khaliq Y, Piper KE, Rouse MS, Patel R. Bacteremia due to viridans group Streptococci with diminished susceptibility to levofloxacin among neutropenic patients receiving levofloxacin prophylaxis. Clin Infect Dis. 2002;34:1469-1474. doi:10.1086/340352
- 119. Zervos MJ, Hershberger E, Nicolau DP, et al. Relationship between fluoroquinolone use and changes in susceptibility to fluoroquinolones of selected pathogens in 10 United States teaching hospitals, 1991-2000. Clin Infect Dis. 2003;37:1643-1648. doi:10.1086/379709

- 120. Kern WV, Steib-Bauert M, de With K, et al. Fluoroquinolone consumption and resistance in haematology-oncology pa- tients: ecological analysis in two university hospitals 1999- 2002. *J Antimicrob Chemother*. 2005;55:57-60. doi:10.1093/jac/dkh510
- 121. Mikulska M, Averbuch D, Tissot F, et al. Fluoroquinolone prophylaxis in haematological cancer patients with neutropenia: ECIL critical appraisal of previous guidelines. *J Infect*. 2018;76: 20-37. doi:10.1016/j.jinf.2017.10.009
- 122. Ramasamy K. Reducing infection-related morbidity and mortality in patients with myeloma. Lancet Oncol. 2019;20:1633-1635. doi:10.1016/S1470-2045(19)30649-7
- 123. Slavin MA, Worth LJ, Seymour JF, Thursky KA. Better sepsis management rather than fluoroquinolone prophylaxis for patients with cancer-related immuno- suppression. *J Clin Oncol.* 2019;37:1139-1140. doi:10.1200/JCO.18.01474
- 124. Verlinden A, Schroyens WA, Gadisseur AP. Clinical and microbiological impact of long-term discontinuation of fluoroquinolone prophylaxis in haematological patients with prolonged profound neutropenia. *Eur J Haematol.* 2021;107:377-379. doi:10.1111/eih.13670
- 125. Centers for Disease Control and Prevention. Preventing Infections in Cancer Patients.

 Accessed November 11, 2020. cdc.gov/cancer/preventinfections/index.htm
- 126. American Cancer Society. Infections in People With Cancer. Accessed November 11, 2020. cancer.org/treatment/treatments -and -side-effects/physical-side-effects/low-blood count s/infections. html
- 127. National Comprehensive Cancer Network. Prevention and Treatment of Cancer-Related Infections. Accessed February 26, 2021, nccn.org/professionals/physician_gls/default.aspx
- 128. Montassier E, Gastinne T, Vangay P, et al. Chemotherapy-driven dysbiosis in the intestinal microbiome. Aliment Pharmacol Ther. 2015;42:515-528. doi:10.1111/ apt.13302
- 129. Dethlefsen L, Huse S, Sogin ML, Relman DA. The pervasive effects of an antibiotic on the human gut microbiota, as revealed by deep 16S rRNA sequencing. *PLoS Biol.* 2008;6:e280. doi:10.1371/journal.pbio.0060280
- 130. Dethlefsen L, Relman DA. Incomplete recovery and individualized responses of the human distal gut microbiota to repeated antibiotic perturbation. *Proc Natl Acad Sci USA*. 2011;108(suppl 1):4554- 4561. doi:10.1073/pnas.10000 87107
- 131. Pilmis B, Le Monnier A, Zahar JR. Gut microbiota, antibiotic therapy and antimicrobial resistance: a narrative review. *Microorganisms*. 2020;8:269. doi:10.3390/microorganisms.020269
- 132. Holler E, Butzhammer P, Schmid K, et al. Metagenomic analysis of the stool microbiome in patients receiving allogeneic stem cell transplantation: loss of diversity is associated with use of systemic antibiotics and more pronounced in gastrointestinal graft- versus- host disease. *Biol Blood Marrow Transplant*. 2014;20:640-645. doi:10.1016/j.bbmt.2014.01.030
- 133. Petrelli F, Iaculli A, Signorelli D, et al. Survival of patients treated with antibiotics and immunotherapy for cancer: a systematic review and meta- analysis. *J Clin Med.* 2020;9:1458. doi:10.3390/icm90.51458
- 134. Kim SG, Becattini S, Moody TU, et al. Microbiota- derived lantibiotic restores resistance against vancomycin-resistant Enterococcus. *Nature*. 2019;572:665-669. doi:10.1038/s4158 6-019-1501-z
- 135. Koh AY. Potential for monitoring gut microbiota for diagnosing infections and graft- versus- host disease in cancer and stem cell transplant patients. *Clin Chem.* 2017;63:1685- 1694. doi:10.1373/clinc hem.2016.259499
- 136. Salgia NJ, Bergerot PG, Maia MC, et al. Stool microbiome profiling of patients with metastatic renal cell carcinoma receiving anti- PD-1 immune checkpoint inhibitors. Eur Urol. 2020;78:498-502. doi:10.1016/i.eururo.2020.07.011
- 137. Willems RJ, Top J, van Santen M, et al. Global spread of vancomycin-resistant Enterococcus faecium from distinct nosocomial genetic complex. *Emerg Infect Dis.* 2005;11:821-828. doi:10.3201/1106.041204
- 138. Arcilla MS, van Hattem JM, Haverkate MR, et al. Import and spread of extendedspectrum beta- lactamase- producing Enterobacteriaceae by international travellers (COMBAT study): a prospective, multicentre cohort study. *Lancet Infect Dis.* 2017;17:78-85. doi:10.1016/S1473 - 3099(16)30319 - X
- 139. Frost I, Van Boeckel TP, Pires J, Craig J, Laxminarayan R. Global geographic trends in antimicrobial resistance: the role of international travel. *J Travel Med.* 2019;26;taz036. doi:10.1093/itm/taz036
- 140. Carling P, Fung T, Killion A, Terrin N, Barza M. Favorable impact of a multidisciplinary antibiotic management program conducted during 7 years. *Infect Control Hosp Epidemiol.* 2003;24:699-706. doi:10.1086/502278
- 141. Schentag JJ, Ballow CH, Fritz AL, et al. Changes in antimicrobial agent usage resulting from interactions among clinical pharmacy, the infectious disease division, and the microbiology laboratory. *Diagn Microbiol Infect Dis.* 1993;16:255-264. doi:10.1016/073 2-8893(93)90119 r
- 142. Ansari F, Gray K, Nathwani D, et al. Outcomes of an intervention to improve hospital antibiotic prescribing: interrupted time series with segmented regression analysis. *J Antimicrob Chemother.* 2003;52:842-848. doi:10.1093/jac/dkg459
- 143. Cole KA, Rivard KR, Dumkow LE. Antimicrobial stewardship interventions to combat antibiotic resistance: an update on targeted strategies. Curr Infect Dis Rep. 2019;21:33.

- doi:10.1007/s1190 8-019-0689-2
- 144. Klepser ME, Adams AJ, Klepser DG. Antimicrobial stewardship in outpatient settings: leveraging innovative physician-pharmacist collaborations to reduce antibiotic resistance. *Health Secur.* 2015;13:166-173. doi:10.1089/hs.2014.0083
- 145. Zhang ZG, Chen F, Ou Y. Impact of an antimicrobial stewardship programme on antibiotic usage and resistance in a tertiary hospital in China. *J Clin Pharm Ther.* 2017;42:579-584. doi:10.1111/jcpt.12544
- 146. Petrelli F, Ghidini M, Ghidini A, et al. Use of antibiotics and risk of cancer: a systematic review and meta- analysis of observational studies. *Cancers* (Basel). 2019;11:1174. doi:10.3390/cancers11081174
- 147. Peyrony O, Gerlier C, Barla I, et al. Antibiotic prescribing and outcomes in cancer patients with febrile neutropenia in the emergency department. PLoS One. 2020;15:e0229828. doi:10.1371/journal.pone.0229828
- 148. Spellberg B. The new antibiotic mantra— "shorter is better." JAMA Intern Med. 2016;176:1254- 1255. doi:10.1001/jamai ntern med.2016.3646
- 149. Koenig C, Schneider C, Morgan JE, Ammann RA, Sung L, Phillips B. Association of time to antibiotics and clinical outcomes in patients with fever and neutropenia during chemotherapy for cancer: a systematic review. Support Care Cancer. 2020;28:1369-1383. doi:10.1007/s0052 0-019-04961 - 4
- 150. Kim NH, Koo HL, Choe PG, et al. Inappropriate continued empirical vancomycin use in a hospital with a high prevalence of methicillin- resistant Staphylococcus aureus.

 Antimicrob Agents Chemother. 2015;59:811-817. doi:10.1128/ AAC.04523 14
- 151. Freifeld AG, Bow EJ, Sepkowitz KA, et al. Clinical practice guideline for the use of antimicrobial agents in neutropenic patients with cancer: 2010 update by the Infectious Diseases Society of America. Clin Infect Dis. 2011;52:e56-93. doi:10.1093/cid/cir073
- 152. Schmidt- Hieber M, Teschner D, Maschmeyer G, Schalk E. Management of febrile neutropenia in the perspective of antimicrobial de- escalation and discontinuation. Expert Rev Anti Infect Ther. 2019;17:983-995. doi:10.1080/14787 210.2019.1573670
- 153. Le Clech L, Talarmin JP, Couturier MA, et al. Early discontinuation of empirical antibacterial therapy in febrile neutropenia: the ANTIBIOSTOP study. *Infect Dis* (Lond). 2018;50:539-549. doi:10.1080/23744.235.2018.1438649
- 154. Heinz WJ, Buchheidt D, Christopeit M, et al. Diagnosis and empirical treatment of fever of unknown origin (FUO) in adult neutropenic patients: guidelines of the Infectious Diseases Working Party (AGIHO) of the German Society of Hematology and Medical Oncology (DGHO). Ann Hematol. 2017;96:1775-1792. doi:10.1007/s00277-017-3098-2
- 155. Rearigh L, Stohs E, Freifeld A, Zimmer A. De- escalation of empiric broad spectrum antibiotics in hematopoietic stem cell transplant recipients with febrile neutropenia. *Ann Hematol.* 2020:99:1917-1924. doi:10.1007/s00277-020-04132-0
- 156. Aguilar- Guisado M, Espigado I, Martin- Pena A, et al. Optimisation of empirical antimicrobial therapy in patients with haematological malignancies and febrile neutropenia (How Long Study): an open-label, randomised, con- trolled phase 4 trial. Lancet Haematol. 2017;4:e573-e583. doi:10.1016/S2352-3026(17)30211-9
- 157. la Martire G, Robin C, Oubaya N, et al. De-escalation and discontinuation strategies in high-risk neutropenic patients: an interrupted time series analyses of antimicrobial consumption and impact on outcome. *Eur J Clin Microbiol Infect Dis.* 2018;37:1931-1940. doi:10.1007/s10096-018-3328-1
- 158. Puerta- Alcalde P, Cardozo C, Suarez- Lledo M, et al. Current time- to- positivity of blood cultures in febrile neutropenia: a tool to be used in stewardship de- escalation strategies. Clin Microbiol Infect. 2019;25:447- 453. doi:10.1016/j.cmi.2018.07.026
- 159. Byun JM, Jeong DH. Antibiotic prophylaxis for gynecologic cancer surgery. *Taiwan J Obstet Gynecol*. 2020;59:514-519. doi:10.1016/j.tjog.2020.05.008
- 160. Gerding DN. The search for good antimicrobial stewardship. *Jt Comm J Qual Improv.* 2001;27:403-404. doi:10.1016/s1070-3241(01)27034-5
- 161. Doron S, Davidson LE. Antimicrobial stewardship. Mayo Clin Proc. 2011;86:1113-1123. doi:10.4065/mcp.2011.0358
- 162. Gudiol C, Carratala J. Antibiotic resistance in cancer patients. Expert Rev Anti Infect Ther. 2014;12:1003-1016. doi:10.1586/14787 210.2014.920253
- 163. Wolf J, Margolis E. Effect of antimicrobial stewardship on outcomes in patients with cancer or undergoing hematopoietic stem cell transplantation. *Clin Infect Dis.* 2020;71:968-970. doi:10.1093/cid/ ciz903
- 164. Rosa RG, Goldani LZ, dos Santos RP. Association between adherence to an antimicrobial stewardship program and mortality among hospitalised cancer patients with febrile neutropaenia: a prospective cohort study. BMC Infect Dis. 2014;14:286. doi:10.1186/1471-2334-14-286
- 165. Pillinger KE, Bouchard J, Withers ST, et al. Inpatient antibiotic stewardship interventions in the adult oncology and hematopoietic stem cell transplant population: a review of the literature. Ann Pharmacother. 2020;54:594-610. doi:10.1177/10600 28019 890886
- 166. Bantar C, Sartori B, Vesco E, et al. A hospitalwide intervention program to optimize the quality of antibiotic use: impact on prescribing practice, antibiotic consumption, cost savings, and bacterial resistance. Clin Infect Dis. 2003;37:180-186. doi:10.1086/375818
- 167. Rahal JJ, Urban C, Horn D, et al. Class restriction of cephalosporin use to control total cephalosporin resistance in nosocomial Klebsiella. *JAMA*. 1998;280:1233-1237. doi:10.1001/jama.280.14.1233

- 168. Cook PP, Catrou P, Gooch M, Holbert D. Effect of reduction in ciprofloxacin use on prevalence of meticillin-resistant Staphylococcus aureus rates within individual units of a tertiary care hospital. *J Hosp Infect*. 2006;64:348-351.
- doi:10.1016/j.jhin.2006.06.033
- 169. Raymond DP, Pelletier SJ, Crabtree TD, et al. Impact of a rotating empiric antibiotic schedule on infectious mortality in an intensive care unit. Crit Care Med. 2001;29:1101-1108. doi:10.1097/00003 246-20010 6000-00001
- 170. Timbrook TT, Hurst JM, Bosso JA. Impact of an antimicrobial stewardship program on antimicrobial utilization, bacterial susceptibilities, and financial expenditures at an academic medical center. *Hosp Pharm.* 2016;51:703-711.
- doi:10.1310/hpi51 09- 703
- 171. Doernberg SB, Abbo LM, Burdette SD, et al. Essential resources and strategies for antibiotic stewardship programs in the acute care setting. *Clin Infect Dis.* 2018;67:1168-1174. doi:10.1093/cid/ciy255
- 172. Centers for Disease Control and Prevention. Core Elements of Outpatient Antibiotic Stewardship. Accessed May 24, 2020. cdc.gov/antibiotic use/core-elements/outpatient. html
- 173. Marcelin JR, Chung P, Van Schooneveld TC. Antimicrobial stewardship in the outpatient setting: a review and proposed framework. *Infect Control Hosp Epidemiol*. 2020;41:833-840. doi:10.1017/ice.2020.94
- 174. US Food and Drug Administration. 2019 Summary Report on Antimicrobials Sold or Distributed for Use in Food- Producing Animals. Accessed May 24, 2021. fda. gov/anima I-veterinary/ cvm- updates/ fda- releases-annual- summary-report- antimicrobials- sold-or- distributed- 2019- use- food- producing
- 175. Martin MJ, Thottathil SE, Newman TB. Antibiotics overuse in animal agriculture: a call to action for health care providers. *Am J Public Health*. 2015;105:2409-2410. doi:10.2105/AJPH.2015.302870
- 176. Landers TF, Cohen B, Wittum TE, Larson EL. A review of antibiotic use in food animals: perspective, policy, and potential. *Public Health Rep.* 2012;127:4-22.
- doi:10.1177/00333 54912 12700103
- 177. Angulo FJ, Nargund VN, Chiller TC. Evidence of an association between use of anti-microbial agents in food animals and anti-microbial resistance among bacteria isolated from humans and the human health consequences of such resistance. *J Vet Med B Infect Dis Vet Public Health*. 2004;51:374-379. doi:10.1111/j.1439-0450.2004.00789.x
- 178. Centers for Disease Control and Prevention. Food and Food Animals. Accessed May 24, 2021. cdc.gov/drugr esist ance/food.html
- 179. Chen Y, Hammer EE, Richards VP. Phylogenetic signature of lateral exchange of genes for antibiotic production and resistance among bacteria highlights a pattern of global transmission of pathogens between humans and livestock. Mol Phylogenet Evol. 2018;125:255-264. doi:10.1016/j.ympev.2018.03.034
- 180. Thacker SB, Berkelman RL. Public health surveillance in the United States. *Epidemiol Rev.* 1988;10:164-190. doi:10.1093/oxfor djour nals.epirev.a036021
- 181. Simonsen GS. Antimicrobial resistance surveillance in Europe and beyond. *Euro Surveill.* 2018;23:1800560. doi:10.2807/1560-7917.ES.2018.23.42. 1800560
- 182. Perez F, Villegas MV. The role of surveillance systems in confronting the global crisis of antibiotic-resistant bacteria. Curr Opin Infect Dis. 2015;28:375-383. doi:10.1097/ QCO.0000 00000 000182
- 183. Tsutsui A, Suzuki S. Japan Nosocomial Infections Surveillance (JANIS): a model of sustainable national antimicrobial resistance surveillance based on hospital diagnostic microbiology laboratories. BMC Health Serv Res. 2018;18:799. doi:10.1186/s12913-018-3604-x
- 184. Johnson AP. Surveillance of antibiotic resistance. Philos Trans R Soc Lond B Biol Sci. 2015;370:20140080. doi:10.1098/rstb.2014.0080
- $185. Kadri SS, Boucher HW. U.S. Efforts to curb antibiotic resistance—are we saving lives? \\N Engl J Med. 2020; 383:806-808. doi:10.1056/NEJMp 2004743$
- 186. Hidron AI, Edwards JR, Patel J, et al. NHSN annual update: antimicrobial-resistant pathogens associated with healthcare- associated infections: annual summary of data reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2006-2007. Infect Control Hosp Epidemiol. 2008;29:996-1011. doi:10.1086/591861
- 187. Lake JG, Weiner LM, Milstone AM, Saiman L, Magill SS, See I. Pathogen distribution and antimicrobial resistance among pediatric healthcare- associated infections reported to the National Healthcare Safety Network, 2011-2014. *Infect Control Hosp Epidemiol*. 2018;39:1-11. doi:10.1017/ice.2017.236
- 188. Sievert DM, Ricks P, Edwards JR, et al. Antimicrobial- resistant pathogens associated with healthcare-associated infections: summary of data reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2009-2010. Infect Control Hosp Epidemiol. 2013;34:1-14. doi:10.1086/668770
- 189. Weiner LM, Webb AK, Limbago B, et al. Antimicrobial-resistant pathogens associated with healthcare-associated infections: summary of data reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2011-2014. Infect Control Hosp Epidemiol. 2016;37:1288-1301. doi:10.1017/ice.2016.174
- 190. Alm RA, Gallant K. Innovation in antimicrobial resistance: the CARB-X perspective. ACS Infect Dis. 2020;6:1317-1322. doi:10.1021/acsin fecdis.0c00026
- $191.\,\mathsf{Rex}\,\mathsf{JH},\mathsf{Fernandez}\,\mathsf{Lynch}\,\mathsf{H},\mathsf{Cohen}\,\mathsf{IG},\mathsf{Darrow}\,\mathsf{JJ},\mathsf{Outterson}\,\mathsf{K}.\,\mathsf{Designing}$

- development programs for non-traditional antibacterial agents. *Nat Commun.* 2019;10:3416. doi:10.1038/s4146 7-019-11303 9
- 192. Rex JH, Outterson K. Antibiotic reimbursement in a model delinked from sales: a benchmark-based worldwide approach. *Lancet Infect Dis*. 2016;16:500-505. doi:10.1016/51473-3099/15100500-9
- 193. Ardal Cm Findlay D, Savic M, et al. Revitalizing the Antibiotic Pipeline: Stimulating Innovation While Driving Sustainable Use and Global Access. Final Report. DRIVE-AB; 2018. Accessed February 21, 2021. drive ab.eu/wp- content/uploads/2018/01/DRIVE AB-Final Report Jan 2018. ddf
- 194. Presidential Advisory Council on Combating Antibiotic- Resistant Bacteria (PACCARB). Recommendations for Incentivizing the Development of Vaccines, Diagnostics, and Therapeutics to Combat Antibiotic-Resistance. PACCARB; 2017. Accessed February 21, 2021. hhs.gov/ web/508/sites/ defau lt/files/ pacca rb- final incentives report- sept- 2017.pdf
- 195. O'Neill J. Review on Antimicrobial Resistance. Tackling Drug-Resistant Infections Globally: Final Report and Recommendations. *Review on Antimicrobial Resistance*; 2016. Accessed February 21, 2021. amr-amr review.org/ sites/ defau lt/files/ 160518_Final %20pap er with%20cov er.pdf
- 196. Towse A, Hoyle CK, Goodall J, Hirsch M, Mestre-Ferrandiz J, Rex JH. Time for a change in how new antibiotics are reimbursed: development of an insurance framework for funding new antibiotics based on a policy of risk mitigation. *Health Policy*. 2017;121:1025-1030. doi:10.1016/j.healt hpol.2017.07.011
- 197. Dutescu IA, Hillier SA. Encouraging the development of new antibiotics: are financial incentives the right way forward? A systematic review and case study. *Infect Drug Resist.* 2021;14:415-434. doi:10.2147/IDR.S287792
- 198. Spaulding CN, Klein RD, Schreiber HLT, Janetka JW, Hultgren SJ. Precision antimicrobial therapeutics: the path of least resistance? NPJ Biofilms Microbiomes. 2018;4:4. doi:10.1038/s41522-018-0048-3
- 199. Karam G, Chastre J, Wilcox MH, Vincent JL. Antibiotic strategies in the era of multidrug resistance. Crit Care. 2016;20:136. doi:10.1186/s13054-016-1320-7
- 200. Boucher HW, Ambrose PG, Chambers HF, et al. White paper: developing antimicrobial drugs for resistant pathogens, narrow-spectrum indications, and unmet needs. J Infect Dis. 2017;216:228-236. doi:10.1093/infdis/lix211
- Melander RJ, Zurawski DV, Melander C. Narrow-spectrum antibacterial agents. Medchemcomm. 2018;9:12-21. doi:10.1039/C7MD0 0528H
- 202. Schooley RT, Biswas B, Gill JJ, et al. Development and use of personalized bacteriophage- based therapeutic cocktails to treat a patient with a disseminated resistant Acinetobacter baumannii infection. Antimicrob Agents Chemother. 2017;61:e00954-17. doi:10.1128/AAC.00954-17
- 203. Regeimbal JM, Jacobs AC, Corey BW, et al. Personalized therapeutic cocktail of wild environmental phages rescues mice from Acinetobacter baumannii wound infections. Antimicrob Agents Chemother. 2016;60:5806-5816. doi:10.1128/AAC.02877-15
- 204. Nielsen TB, Pantapalangkoor P, Luna BM, et al. Monoclonal antibody protects against Acinetobacter baumannii infection by enhancing bacterial clearance and evading sepsis. J Infect Dis. 2017;216:489-501. doi:10.1093/infdis/jix315
- 205. Cotter PD, Ross RP, Hill C. Bacteriocins—a viable alternative to antibiotics? Nat Rev Microbiol. 2013;11:95- 105. doi:10.1038/nrmic ro2937
- 206. Ghequire MG, Dingemans J, Pirnay JP, De Vos D, Cornelis P, De Mot R. O serotypeindependent susceptibility of Pseudomonas aeruginosa to lectin-like pyocins. *Microbiologyopen*. 2014;3:875-884. doi:10.1002/mbo3.210
- $207. \, Howard \, JJ, Sturge \, CR, \, Moustafa \, DA, et al. \, Inhibition \, of \, Pseudomonas \, aeruginosa \, \\ by \, peptide-conjugated \, phosphorodiami- \, date \, morpholino \, oligomers. \, \textit{Antimicrob Agents}$

- Chemother. 2017;61:e01938-16. doi:10.1128/AAC.01938-16
- 208. Moustafa DA, Wu AW, Zamora D, et al. Peptide-conjugated phosphorodi-amidate morpholino oligomers retain activity against multidrug-resistant Pseudomonas aeruginosa in vitro and in vivo. mBio. 2021;12:e02411- 20. doi:10.1128/mBio.02411-20
- 209. Geller BL, Li L, Martinez F, et al. Morpholino oligomers tested in vitro, in biofilm and in vivo against multidrug-resistant Klebsiella pneumoniae. *J Antimicrob Chemother.* 2018;73:1611- 1619. doi:10.1093/jac/dky058
- 210. Sulakvelidze A, Alavidze Z, Morris JG Jr. Bacteriophage therapy. Antimicrob Agents Chemother. 2001;45:649-659. doi:10.1128/AAC.45.3.649-659.2001
- 211. Drulis- Kawa Z, Majkowska-Skrobek G, Maciejewska B, Delattre AS, Lavigne R. Learning from bacteriophages— advantages and limitations of phage and phage- encoded protein applications. Curr Protein Pept Sci. 2012;13:699-722. doi:10.2174/13892.03128.04871193
- 212. Zurawski DV, McLendon MK. Monoclonal antibodies as an antibacterial approach against bacterial pathogens. *Antibiotics* (Basel). 2020;9:155. doi:10.3390/antibiotic s9040155
- 213. Motley MP, Banerjee K, Fries BC. Monoclonal antibody-based therapies for bacterial infections. *Curr Opin Infect Dis.* 2019;32:210-216. doi:10.1097/
- QCO.00000 00000 000539
- 214. Joerger RD. Alternatives to antibiotics: bacteriocins, antimicrobial peptides and bacteriophages. *Poult Sci.* 2003;82:640-647. doi:10.1093/ps/82.4.640
- 215. Yang SC, Lin CH, Sung CT, Fang JY. Antibacterial activities of bacteriocins: application in foods and pharmaceuticals. Front Microbiol. 2014;5:241. doi:10.3389/ fmicb.2014.00241
- 216. Shelburne SA, Kim J, Munita JM, et al. Whole-genome sequencing accurately identifies resistance to extended-spectrum beta-lactams for major Gram-negative bacterial pathogens. Clin Infect Dis. 2017;65:738-745. doi:10.1093/cid/cix417
- 217. Li X, Lin J, Hu Y, Zhou J. PARMAP: a pan-genome-based computational framework for predicting antimicrobial resistance. Front Microbiol. 2020;11:578795. doi:10.3389/ fmicb.2020.578795
- 218. Hubbard ATM, Mason J, Roberts P, et al. Piperacillin/tazobactam resistance in a clinical isolate of Escherichia coli due to IS26- mediated amplification of blaTEM- 1B. Nat Commun. 2020;11:4915. doi:10.1038/s41467-020-18668-2
- 219. Maurer FP, Christner M, Hentschke M, Rohde H. Advances in rapid identification and susceptibility testing of bacteria in the clinical microbiology laboratory: implications for patient care and antimicrobial stewardship programs. *Infect Dis Rep.* 2017;9:6839. doi:10.4081/idr.2017.6839
- 220. Maxson T, Mitchell DA. Targeted treatment for bacterial infections: prospects for pathogen-specific antibiotics coupled with rapid diagnostics. *Tetrahedron*. 2016;72:3609-3624. doi:10.1016/j.tet.2015.09.069
- 221. Trevas D, Caliendo AM, Hanson K, Levy J, Ginocchio CC. Diagnostic tests can stem the threat of antimicrobial resistance: infectious disease professionals can help. Clin Infect Dis. 2021;72:e893- e900. doi:10.1093/cid/ ciaa1527
- 222. Tamma PD, Aitken SL, Bonomo RA, Mathers AJ, van Duin D, Clancy CJ. Infectious Diseases Society of America Guidance on the treatment of extended-spectrum beta- lactamase producing Enterobacterales (ESBL- E), carbapenem-resistant Enterobacterales (CRE), and Pseudomonas aeruginosa with difficult- to- treat resistance (DTR- P. aeruginosa). Clin Infect Dis. 2021;72:e169-e183. doi:10.1093/cid/ciaa1478
- 223. Baselga J, Bhardwaj N, Cantley LC, et al. AACR Cancer Progress Report 2015. Clin Cancer Res. 2015;21(19 suppl):S1-S128. doi:10.1158/1078-0432.CCR-15-1846